0957-4166(95)00284-7

# Enzymatic Synthesis of (S)-(-)-1-(2-Thienyl)propyl Acetate

# Suk-Ku Kang,\* Jae-Ho Jeon, Tokutaro Yamaguchi, Jae-Sun Kim, and Byoung-Seob Ko

Department of Chemistry, Sung Kyun Kwan University, Natural Science Campus, Suwon 440-746, Korea

**Abstract:** ( $\pm$ )-1-(2-Thienyl)propyl acetate was resolved by PCL (*Pseudomonas cepacia* lipase) catalyzed hydrolysis to afford (S)-(-)-1-(2-thienyl) propyl acetate in >99% e.e.. (S)-(-)-1-(2-Thienyl) propyl acetate thus obtained was transformed to (S)-(+)-3-octanol, the alarm pheromone of ants, *Crematogaster castanea* and *liengmei*.

Enzyme-catalyzed kinetic resolution of racemic secondary alcohols to prepare enantiomerically pure secondary alcohols has become an important method in organic synthesis. No enzymatic resolution of 2-thienyl alcohols or acetates has been reported although the preparation of optically active 2-thienyl alcohols by chemical resolution using Sharpless reagent was known. In connection with our project on the synthesis of new chiral building blocks for the synthesis of optically active insect pheromones, we report here our results on the lipase-catalyzed preparation of (S)-(-)-1-(2-thienyl)propyl accetate (S)-(-)-1 (Scheme 1).

Scheme 1

The results of enzymatic hydrolysis of  $(\pm)$ -1 catalyzed by PCL are summarized in Table 1. Of the lipases tested,<sup>3</sup> PCL exhibited the highest enantioselectivity. A notable feature of this transformation is that the enantioselectivity is highly dependent on temperature. The system of 0.1 M acetone-phosphate buffer was the most effective for enhancing the enantioselectivity. When the reaction was run at 22 °C and stopped at 60% conversion, the % e.e. of the remaining acetate was 89% e.e. (entry 1 in Table 1). For 30% conversion at 22°C, the acetate was obtained in 82% e.e. (entry 2). The enantiomer (S)-(-)-1 of >99% e.e. was obtained at 18 °C at 60% conversion (entry 3). In this conversion, the specific rotation of (S)-(-)-1 was  $[\alpha]_D^{25}$  -157.8 (C 0.65, CHCl<sub>3</sub>). However at 14 °C for 8 h (30% conversion), the acetate with 73 % e.e. was obtained (entry 4).

The absolute configuration of (S)-1 was confirmed by transformation of (S)-1 to (S)-(+)-3-octanol (S)-4,<sup>4.5</sup> the alarm pheromone of ant, *Crematogaster castanea* and *liengmei* which was shown in Scheme 2. Hydrolysis of (S)-1 with  $K_2CO_3$  in MeOH at room temperature for 0.5 h provided (S)-2,<sup>6</sup> [ $\alpha$ ]<sup>25</sup><sub>D</sub> -25.3 (c 1.6, CHCl<sub>3</sub>) which was indirectly correlated to (R)-2,<sup>5</sup> [ $\alpha$ ]<sup>25</sup><sub>D</sub> +25.9 (c 2.1, CHCl<sub>3</sub>). Lithiation of (S)-2 with n-BuLi followed by methylation with MeI afforded (S)-3,<sup>5</sup> [ $\alpha$ ]<sup>25</sup><sub>D</sub> -29.1 (c 1.2, CHCl<sub>3</sub>) in 66% overall yield.<sup>7</sup> Finally, reductive desulfurization with Ra-Ni of (S)-3 provided (S)-4 (Scheme 2).

Entry	lipase <sup>b</sup> (mg, mmol of substrate	Temp. ℃	Time (h)	Acetate % e.e. <sup>c</sup> (% <sup>d</sup> ) [α] <sub>D</sub> <sup>25 e</sup>	Alcohol % e.e. <sup>c</sup> (% <sup>d</sup> ) [α] <sup>25 c</sup>	Ef
1	PCL (36.7)	22	13	89 (18.8) -144.5	69 (33.3) +19.2	15.9
2	PCL (34.8)	22	6	82 (33.4) -133.2	75(30.7)+21.2	17.5
3	PCL (34.9)	18	20	>99 (25) -157.8	71(57.3)+20.0	30.3
_4	PCL (33.5)	14	8	73 (58) -119.1	75 (32) +20.5	15.3

**Table 1** Results of the Lipase Catalyzed Hydrolysis of  $(\pm)-1^a$ 

Scheme 2

#### Experimental

<sup>1</sup>H-NMR specra were determined in CDCl<sub>3</sub> as solvent using Gemini 200 spectrometer ( <sup>1</sup>H 199.98 MHz). Chemical shifts are given in ppm downfield from tetramethyl silane. IR spectra were obtained from a Nicolet Model 205 FT-IR spectrometer and are reported in cm<sup>-1</sup>. Specific rotation was measured on a Rudolph Autopol polarimeter at 25°C. Mass spectra were taken on a VG Trio 2000 instrument. Elemental Analyses were performed on a EA 1108 (Carlo Erba instrument).

#### (S)-1-(2-Thienyl) propyl acetate [(S)-1]

To a stirred solution of  $(\pm)$ -1 (301 mg, 1.64 mmol) in acetone (4 mL) at 18 °C was added 0.1 M phosphate buffer (pH 7.1, 100 mL), was added PCL (57 mg) at one portion and the reaction mixture was shaken at 18 °C for 20 h in water bath shaker and stopped at 60% conversion of  $(\pm)$ -1 to (R)-2. The enzymatic hydrolysis was monitored using Varian 3700 gas chromatography (Chrom G-HP, 100/120 50 cm x 1/8") [GC conditions: injection temp 120 °C, detection temp; 160 °C; flow rate of carrier N<sub>2</sub> gas 20 psi. The column temperature was maintained at 50 °C for 5 min and increased at a rate of 10 °C/min to 120 °C. The retention time of the alcohol was 7.45 min and that of the acetate was 9.58 min.] The reaction mixture was extracted with ether (100 mL) for three times and then evaporated *in vacuo*. The crude product was separated by SiO<sub>2</sub> column chromatograhy using ethyl acetate/hexanes (1:5) as eluent to afford (S)-1 (76.1 mg, 25%) in >99% e.e. and (R)-2 (133.6 mg, 57%) in 71% e.e.. (S)-1: TLC; SiO<sub>2</sub>, EtOAc/hexanes 1:15, R<sub>5</sub> = 0.60. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  0.95 (t,

<sup>&</sup>lt;sup>a</sup> The compound (±)-1 was prepared from thiophene-2-carboxaldehyde by Grignard addition of EtMgBr followed by acetylation. <sup>b</sup> The enzyme *Pseudomonas cepacia* lipse (PCL) used was supplied by Amano pharmaceutical Co. Ltd.. <sup>c</sup> % e.e. was checked by GC-MS of MTPA esters. <sup>d</sup> The yields are isolated yields. <sup>e</sup> The values in CHCl<sub>3</sub> at 25 °C. <sup>f</sup> See: Chen, C-S.; Fujimoto, Y.; Girdaukas, G; Sih, C. J. *J. Am. Chem. Soc.* 1982, 104, 7294.

3H, J = 8.5 Hz), 1.85-2.00 (m, 2H), 2.05 (s, 3H), 5.95 (t, 1H, J = 6.0 Hz), 6.95 (m, 1H), 7.05 (m, 1H), 7.25 (m, 1H). IR (film) 1745 cm<sup>-1</sup>.MS(m/z) 184(M<sup>+</sup>), 142, 123, 113(base peak), 97, 85. Anal. Calcd. for  $C_9H_{12}O_2S : C$ , 58.7; H, 6.56; S, 17.4. Found: C, 58.3; H, 6.59; S, 17.1. (*R*)-2: TLC; SiO<sub>2</sub>, EtOAc/hexanes 1: 15,  $R_f = 0.36$ . <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.00 (t, 3H, J = 8.5 Hz), 1.90 (m, 2H), 2.05 (s, 3H), 4.89 (t, 1H, J = 6.0 Hz,), 7.00 (d, 1H, J = 3.5 Hz), 7.25 (m, 1H), 7.80 (m, 1H). IR (film) 3360, 2950, 1450 cm<sup>-1</sup>

#### (S)-1-(2-Thienyl) propanol [(S)-2]

To a stirred solution of (*S*)-1 (300 mg, 1.64 mmol) in dry MeOH (5 mL) was added anhydrous  $K_2CO_3$  (0.23 g, 1.63 mmol) in one portion. The reaction mixture was stirred for 0.5 h and then MeOH was evaporated and extracted with ether (30 mL). The ether layer was washed with brine, dried over anhydrous MgSO<sub>4</sub>, and evaporated *in vacuo*. The crude product was separated by  $SiO_2$  chromatograhy using EtOAc/hexanes (1 : 30) as eluent to afford (*S*)-2 (228 mg, 98%). (*S*)-2: TLC;  $SiO_2$ , EtOAc/hexanes 1 : 15,  $R_f = 0.36$ . H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.00 (t, 3H, J = 8.5 Hz), 1.90 (m, 2H), 2.05 (s, 3H), 4.89 (t, 1H, J = 6.0 Hz,), 7.00 (d, 1H, J = 3.5 Hz), 7.25 (m, 1H), 7.80 (m, 1H). IR (film) 3360, 2950, 1450 cm<sup>-1</sup>. MS(m/z) 142(M<sup>+</sup>), 113(base peak), 85. The value of % e.e. was checked by GC-MS analysis (Hewlett-Packard 5890 GC-MS) of (*R*)-MTPA ester (column: HP-1, 25 m × 0.32 mm x 0.52 mm, carrier gas:  $N_2$ , injection temperature: 250 °C, ion source temperature 180 °C). The column temperature was maintained at 180 °C for 5 min and increased at a rate of 5 °C/min to 280 °C. The values of retention times for the MTPA esters were 8.51 min for the (*S*)-isomer and 8.38 min for the (*R*)-isomer.

#### (S)-(-)-1-(5-Methyl-2-Thienyl)propanol [(S)-3]

To a stirred solution of n-BuLi (2.5 M in hexanes, 0.74 mL, 1.9 mmol) and TMEDA (0.28 mL, 1.9 mmol) was added (S)-2 (125 mg, 0.89 mmol) in dry THF (2 mL) at -78 °C. The reaction mixture was stirred for 1 h and warmed to room temperature and then MeI (380 mg, 2.60 mmol) was added slowly and stirred for additional 2 h. The reaction mixture was poured into ice water and extracted with ether. The ether layer was washed with saturated NaCl solution, dried over anhydrous MgSO<sub>4</sub>, and evaporated *in vacuo*. The crude product was separated by SiO<sub>2</sub> chromatograhy using EtOAc/hexanes (1:15, R<sub>f</sub> = 0.36) as eluent to afford (S)-3 (92 mg, 66%). [ $\alpha$ ]<sup>25</sup><sub>D</sub> -29.1 (c 1.2, CHCl<sub>3</sub>). <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  0.95 (t, 3H, J = 8.3 Hz), 1.70-1.95 (m, 2H), 2.15 (bs, 1H), 2.50 (s, 3H), 4.75 (t, 1H), 6.60 (d, 1H, J = 2.8 Hz),6.95 (d, 1H, J = 2.8 Hz). IR (film) 3400 cm <sup>1</sup>. MS(m/z) 156(M<sup>4</sup>) 127(base peak), 99, 65. Anal. Calcd. for C<sub>8</sub>H<sub>12</sub>OS : C, 61.5 ; H. 7.74 ; S, 20.5. Found: C, 61.2 ; H, 7.79 ; S, 20.4.

# (S)-3=Octanol [(S)-4]

To a stirred solution of (S)-3 (120 mg, 0.77 mmol) in MeOH (5 mL) was added Ra-Ni (50% w/w in H<sub>2</sub>O solution) under 1 atm of H<sub>2</sub> atmosphere. The reaction mixture was stirred at room temperature for 1 h and then stirred at room temperature for 1 h and then filtered through silica gel pad. The filtrate was evaporated *in vacuo*. The crude product was separated by SiO<sub>2</sub> chromatograhy using EtOAc/hexanes (1 : 6, R<sub>f</sub> = 0.68) as eluent to afford (S)-4 (77.2 mg, 78%). <sup>1</sup>H NMR (200MHz, CDCl<sub>3</sub>)  $\delta$  0.95 (t, 3H, J = 7.5Hz), 1.20-1.70 (m, 11H), 3.50 (m, 1H). IR(film) 3400, 1450, 1370 cm<sup>-1</sup>.

# Acknowledgments

Financial support by the Korea Research Foundation (1993) by Non Directed Research Fund and Ministry of Education (BSRI-94-3420) is gratefully acknowledged.

### References and Notes

- 1. (a) Santaniello, E.; Ferraboschi, P.; Grisenti, P.; Manzocchi, A. Chem. Rev. 1992, 92, 1071. (b) Poppe, L.; Novak, L.; Selective Biocatalysis. A Synthetic Approach; VCH: Veinheiu, 1992. c) Faber, K. Biotransformations in Organic Chemistry; Springer-Verlag: Berlin, 1992.
- 2. Kitano, Y.; Kunsakae, M.; Kobayashi, Y.; Sato, F. J. Org. Chem. 1989, 54, 994.
- 3. We have tried with PPL(Porcine Pancreas lipase), PFL(Pseudomonas fluorescences lipase), and CRL(Candida rugosa lipase) without success.
- 4. Brand, J. M. J. Chem. Ecol. 1985. 11, 177.
- 5. Fujiwara, M., Mori, K. Agric. Biol. Chem. 1986, 50, 2925.
- 6. Hayashi, M.; Kaneko, T.; Oguni, N. J. Chem. Soc. Perkin. Trans. I. 1991, 25.
- 7. In our hands, enzymatic resolution of (±)-1-(5-methyl-2-thienyl)propyl acetate, (±)-3 was not successful.

(Received in Japan 26 July 1995)